pyhmmer

Release 0.1.4

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Jan 18, 2021

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Cython bindings and Python interface to HMMER3.

CHAPTER

ONE

SETUP

Run pip install pyhmmer in a shell to download the latest release and all its dependencies from PyPi, or have a look at the *Installation page* to find other ways to install pyhmmer.

CHAPTER

TWO

LIBRARY

2.1 Installation

Note: Wheels are provided for Linux x86-64 platforms, but other machines will have to build the wheel from the source distribution. Building pyhmmer involves compiling HMMER3 and Easel, which requires a C compiler to be available.

2.1.1 PyPi

pyhmmer is hosted on GitHub, but the easiest way to install it is to download the latest release from its PyPi repository. It will install all dependencies then install pyhmmer either from a wheel if one is available, or from source after compiling the Cython code :

\$ pip install --user pyhmmer

2.1.2 EMBL Package Registry

You can also install manylinux wheels built from the latest commit that passed the unit tests. Those bleeding-edge releases are available in the GitLab Package Registry hosted on the EMBL git server. Just instruct pip to use an extra index URL as follow:

```
$ pip install --user pyhmmer --extra-index-url https://git.embl.de/api/v4/projects/

$$3638/packages/pypi/simple
```

2.1.3 GitHub + pip

If, for any reason, you prefer to download the library from GitHub, you can clone the repository and install the repository by running (with the admin rights):

\$ pip install --user https://github.com/althonos/pyhmmer/archive/master.zip

Caution: Keep in mind this will install always try to install the latest commit, which may not even build, so consider using a versioned release instead.

2.1.4 GitHub + setuptools

If you do not want to use pip, you can still clone the repository and run the setup.py file manually, although you will need to install the build dependencies (mainly Cython):

```
$ git clone --recursive https://github.com/althonos/pyhmmer
$ cd pyhmmer
$ python setup.py build
# python setup.py install
```

Danger: Installing packages without pip is strongly discouraged, as they can only be uninstalled manually, and may damage your system.

2.2 Examples

2.2.1 Active Site Analysis

This example is adapted from the method used by AntiSMASH to annotate biosynthetic gene clusters. AntiSMASH uses profile HMMs to annotate enzymatic domains in protein sequences. By matching the amino acids in the alignment, it can then predict the product specificity of the enzyme.

In this notebook, we show how to reproduce this kind of analysis, using a PKSI Acyltransferase domain built by the AntiSMASH authors (the HMM in HMMER2 format can be downloaded from their git repository).

References

- Del Vecchio, F., H. Petkovic, S. G. Kendrew, L. Low, B. Wilkinson, R. Lill, J. Cortes, B. A. Rudd, J. Staunton, and P. F. Leadlay. 2003. *Active-site residue, domain and module swaps in modular polyketide synthases.* J Ind. Microbiol Biotechnol 30:489-494.
- Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. *antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences.* Nucleic Acids Res. 2011 Jul:W339-46.

[1]: import pyhmmer pyhmmer.__version__

[1]: '0.1.4'

Loading the HMM

Loading a HMMER profile is done with the pyhmmer.plan7.HMMFile class, which provides an iterator over the HMMs in the file. Since we only use a single HMM, we can simply use next to get the first (and only) pyhmmer. plan7.HMM.

```
[2]: with pyhmmer.plan7.HMMFile("data/hmms/txt/PKSI-AT.hmm") as hmm_file:
    hmm = next(hmm_file)
```

Building digitized sequences

Easel provides the code necessary to load sequences from files in common biological formats, such as GenBank or FASTA. These utilities are wrapped by the pyhmmer.easel.SequenceFile, which provides an iterator over the sequences in the file. Note that SequenceFile tries to guess the format by default, but you can force a particular format with the format keyword argument.

```
[3]: with pyhmmer.easel.SequenceFile("data/seqs/PKSI.faa") as seq_file:
        sequences = [ seq.digitize(hmm.alphabet) for seq in seq_file ]
```

Note

The C interface of Easel allows storing a sequence in two different modes: in *text* mode, where the sequence letters are represented as individual characters (e.g. "A" or "Y"), and digital mode, where sequence letters are encoded as digits. To make Python programs clearer, and to allow static typecheck of the storage mode, we provide two separate classes, TextSequence and DigitalSequence, that represent a sequence stored in either of these modes.

SequenceFile yields sequences in text mode, but HMMER expects sequences in digital mode, so we must digitize them. This requires the sequence alphabet to be known, but we can just use the Alphabet instance stored in the alphabet attribute of hmm.

Running a search pipeline

With the sequences and the HMM ready, we can finally run the search pipeline: it has to be initialized with an Alphabet instance, so that the Plan7 background model can be configured accordingly. Then, we run the pipeline in search mode, providing it one HMM, and several sequences. This method returns a TopHits instance that is already sorted and thresholded.

```
[4]: pipeline = pyhmmer.plan7.Pipeline(hmm.alphabet)
    hits = pipeline.search(hmm, sequences)
```

Rendering the alignments

Domain instances store all the required information to report results in their alignment attribute. We can show the alignment between a HMM and a sequence like hmmsearch would as follow (using the first domain of the first hit as an example):

```
[5]: ali = hits[0].domains[0].alignment
              print(" "*3, ali.target_name.decode())
              print("{:3}".format(ali.hmm_from), ali.hmm_sequence, "{:3}".format(ali.hmm_to))
              print(" "*3, ali.identity_sequence)
              print("{:3}".format(ali.target_from), ali.target_sequence, "{:3}".format(ali.target_
                \rightarrowto))
              print(" "*3, ali.hmm_name.decode())
                          sp|Q9ZGI5|PIKA1_STRVZ
                    1
                \rightarrow \texttt{lpggGaMlaVraseeevrelLapyggrlsiAAvNGPrsvVvSGdaeaieallaeLeaqGirarrLkVsHAFHSplMepmldeleevlagitpraPriProduction and a statement of the state
                ⇔308
                           +FpGQG+Q+aGMG eL++++ VF++a+ +C+aaL+p++++sL +v ++ +q
                                                                                                                                                                                                              a+ L++++++OP+ FAv+++LAr
                →W+ Gv+P+AV+GHS+GE++AA+vAG+lSL+DA+r+V R++ ++a l+q+G+Ml+ ++se+ v e+La+++ +ls+AAvNGP.
                →++VvSGd+ +ie+l++++ea_G+rar_++V++A+HS+++e_+_el+evlag++p+aPr+P++S++_G+w+t+
(continues on next page)
                →+ld++YW+r+lR+ V Fa+++etL+ + G+t+F+Ev++hpvLt ++ t
                                                                                                                                                                                                                        + la+Lrr+
```

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```
635 VFPGQGTQWAGMGAELLDSSAVFAAAMAECEAALSPYVDWSLEAVVRQAPG-----

→APTLERVDVVQPVTFAVMVSLARVWQHHGVTPQAVVGHSQGEIAAAYVAGALSLDDAARVVTLRSKSIAAhLAGKGGMLSLALSEDAVLERLAGFD-

→GLSVAAVNGPTATVVSGDPVQIEELARACEADGVRARVIPVDYASHSRQVEIIESELAEVLAGLSPQAPRVPFFSTLEGAWITE-

→PVLDGGYWYRNLRHRVGFAPAVETLATDEGFTHFVEVSAHPVLTMALPGTV----TGLATLRRD 925

PKS-AT.tcoffee
```

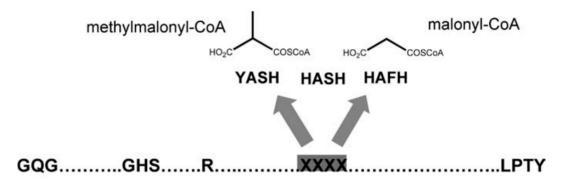
You may also want to see where the domains are located in the input sequence; using the DNA feature viewer developed by the Edinburgh Genome Foundry, we can build a summary graph aligning the protein sequences to the same reference axis:

```
[6]: from dna_features_viewer import GraphicFeature, GraphicRecord
     import matplotlib.pyplot as plt
     # create an index so we can retrieve a Sequence from its name
     seq_index = { seq.name:seq for seq in sequences }
     fig, axes = plt.subplots(nrows=len(hits), figsize=(16, 6), sharex=True)
      for ax, hit in zip(axes, hits):
          # add one feature per domain
          features = [
               GraphicFeature(start=d.alignment.target_from-1, end=d.alignment.target_to)
               for d in hit.domains
          1
          length = len(seq_index[hit.name])
          desc = seq_index[hit.name].description.decode()
          # render the feature records
          record = GraphicRecord(sequence_length=length, features=features)
          record.plot(ax=ax)
          ax.set_title(desc)
      # make sure everything fits in the final graph!
     fig.tight_layout()
     /home/docs/checkouts/readthedocs.org/user_builds/pyhmmer/envs/v0.1.4/lib/python3.7/
      →site-packages/traitlets/traitlets.py:3036: FutureWarning: --rc={'figure.dpi': 96}_
      →for dict-traits is deprecated in traitlets 5.0. You can pass --rc <key=value> ....
      \rightarrow multiple times to add items to a dict.
        FutureWarning,
                     Narbonolide/10-deoxymethynolide synthase PikA1, modules 1 and 2 OS=Streptomyces venezuelae OX=54571 GN=pikAl PE=1 SV=1
                       Narbonolide/10-deoxymethynolide synthase PikA4, module 6 OS=Streptomyces venezuelae OX=54571 GN=pikAIV PE=1 SV=1
                         Mycolipanoate synthase OS=Mycobacterium tuberculosis (strain ATCC 25618 / H37Rv) OX=83332 GN=msl3 PE=1 SV=2
                        Lovastatin nonaketide synthase, polyketide synthase component OS=Aspergillus terreus OX=33178 GN=lovB PE=1 SV=1
                                   Poliketide synthase 37 OS=Dictyostelium discoideum OX=44689 GN=StIB PE=1 SV=1
                300
                                               1.200
                                                          1.500
                                                                    1.800
                                                                               2.100
                                                                                          2.400
                                                                                                     2.700
```

Checking individual positions for catalytic activity

First let's define a function to iterate over an alignement; this will come in handy later. This function yields the position in the alignment (using the HMM coordinates) and the aligned amino acid, skipping over gaps in the HMM sequence.

Now, for the final step, we want to check for the specificity of the enzyme domains; Del Vecchio *et al.* have identified two amino acids in the acyltransferase domain that once muted will decide of the enzyme specificity for either malonyl-CoA or methylmalonyl-CoA:



For this, we need to check the alignment produced by HMMER, and verify the residues of the catalytic site correspond to the ones expected by the authors. We use the function we defined previously, first to check the core amino acids are not muted, and then to check the specificity of the two remaining residues.

```
= [ 93, 94, 95, 120, 196, 198]
[8]: POSITIONS
                = ['G', 'H', 'S', 'R', 'A', 'H']
    EXPECTED
    SPECIFICITY = [195, 197]
    for hit in hits:
        print("\nIn sequence {!r}:".format(hit.name.decode()))
        for domain in hit.domains:
            aligned = dict(iter_target_match(domain.alignment))
            print ("- Found PKSI-AT domain at positions {:4} to {:4}".format (domain.ali_

→from, domain.ali_to))

            try:
                signature = [ aligned[x] for x in POSITIONS ]
                spec = [ aligned[x] for x in SPECIFICITY ]
            except KeyError:
                print(" -> Domain likely too short")
                continue
            if signature != EXPECTED:
                print(" -> Substrate specificity unknown")
            elif spec == ["H", "F"]:
                print(" -> Malonyl-CoA specific")
            elif spec == ["Y", "S"]:
```

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```
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           print(" -> Methylmalonyl-CoA specific")
        else:
           print(" -> Neither malonyl-CoA nor methylmalonyl-CoA specific")
In sequence 'sp|Q9ZGI5|PIKA1_STRVZ':
- Found PKSI-AT domain at positions 635 to 925
 -> Methylmalonyl-CoA specific
- Found PKSI-AT domain at positions 1651 to 1927
 -> Methylmalonyl-CoA specific
- Found PKSI-AT domain at positions 3181 to 3475
 -> Malonyl-CoA specific
In sequence 'sp|Q9ZGI2|PIKA4_STRVZ':
- Found PKSI-AT domain at positions 563 to 837
 -> Methylmalonyl-CoA specific
In sequence 'sp|A0A089QRB9|MSL3_MYCTU':
- Found PKSI-AT domain at positions 540 to 834
 -> Neither malonyl-CoA nor methylmalonyl-CoA specific
In sequence 'sp|Q9Y8A5|LOVB_ASPTE':
- Found PKSI-AT domain at positions 562 to 585
 -> Domain likely too short
- Found PKSI-AT domain at positions 651 to 854
 -> Neither malonyl-CoA nor methylmalonyl-CoA specific
In sequence 'sp|Q54FI3|STLB_DICDI':
- Found PKSI-AT domain at positions 625 to 726
 -> Domain likely too short
- Found PKSI-AT domain at positions 766 to 838
 -> Domain likely too short
- Found PKSI-AT domain at positions 880 to 944
 -> Domain likely too short
```

2.3 API Reference

2.3.1 Errors

Common errors and status codes for the easel and hmmer modules.

```
exception pyhmmer.errors.AllocationError (MemoryError) A memory error that is caused by an unsuccessful allocation.
```

```
exception pyhmmer.errors.UnexpectedError(RuntimeError)
An unexpected error that happened in the C code.
```

As a user of this library, you should never see this exception being raised. If you do, please open an issue with steps to reproduce on the bug tracker, so that proper error handling can be added to the relevant part of the bindings.

```
exception pyhmmer.errors.EaselError (RuntimeError) An error that was raised from the Easel code.
```

2.3.2 Easel

High-level interface to the Easel C library.

Easel is a library developed by the Eddy/Rivas Lab to facilitate the development of biological software in C. It is used by HMMER and Infernal.

Alphabet

class pyhmmer.easel.Alphabet

A biological alphabet, including additional marker symbols.

This type is used to share an alphabet to several objects in the easel and plan7 modules. Reference counting helps sharing the same instance everywhere, instead of reallocating memory every time an alphabet is needed.

Use the factory class methods to obtain a default *Alphabet* for one of the three biological alphabets:

```
>>> dna = Alphabet.dna()
>>> rna = Alphabet.rna()
>>> aa = Alphabet.amino()
```

amino()

Create a default amino-acid alphabet.

dna()

Create a default DNA alphabet.

rna()

Create a default RNA alphabet.

к

The alphabet size, counting only actual alphabet symbols.

Example

```
>>> Alphabet.dna().K
4
>>> Alphabet.amino().K
20
```

Type int

Kp

The complete alphabet size, including marker symbols.

Example

```
>>> Alphabet.dna().Kp
18
>>> Alphabet.amino().Kp
29
```

Type int

symbols

The symbols composing the alphabet.

Example

```
>>> Alphabet.dna().symbols
'ACGT-RYMKSWHBVDN*~'
>>> Alphabet.rna().symbols
'ACGU-RYMKSWHBVDN*~'
```

Type str

Bitfield

class pyhmmer.easel.Bitfield

A statically sized sequence of booleans stored as a packed bitfield.

A bitfield is instantiated with a fixed length, and all booleans are set to False by default:

```
>>> bitfield = Bitfield(8)
>>> len(bitfield)
8
>>> bitfield[0]
False
```

count()

Count the number occurrences of value in the bitfield.

If no argument is given, counts the number of True occurences.

Example

```
>>> bitfield = Bitfield(8)
>>> bitfield.count(False)
8
>>> bitfield[0] = bitfield[1] = True
>>> bitfield.count()
2
```

toggle()

Switch the value of one single bit.

Example

```
>>> bitfield = Bitfield(8)
>>> bitfield[0]
False
>>> bitfield.toggle(0)
>>> bitfield[0]
True
>>> bitfield.toggle(0)
>>> bitfield.toggle(0)
False
```

KeyHash

```
class pyhmmer.easel.KeyHash
```

A dynamically resized container to store string keys using a hash table.

Multiple Sequence Alignment

class pyhmmer.easel.MSA

An abstract alignment of multiple sequences.

checksum()

Calculate a 32-bit checksum for the multiple sequence alignment.

class pyhmmer.easel.TextMSA(MSA)

A multiple sequence alignement stored in text mode.

copy()

Duplicate the text sequence alignment, and return the copy.

digitize()

Convert the text alignment to a digital alignment using alphabet.

class pyhmmer.easel.DigitalMSA(MSA)

A multiple sequence alignment stored in digital mode.

alphabet

The biological alphabet used to encode this sequence alignment to digits.

Type Alphabet, readonly

copy()

Duplicate the digital sequence alignment, and return the copy.

Sequence

class pyhmmer.easel.Sequence

An abstract biological sequence with some associated metadata.

Easel provides two different mode to store a sequence: text, or digital. In the HMMER code, changing from one mode to another mode is done in place, which allows recycling memory. However, doing so can be confusing since there is no way to know statically the representation of a sequence.

To avoid this, pyhmmer provides two subclasses of the Sequence abstract class to maintain the mode contract: TextSequence and DigitalSequence. Functions expecting sequences in digital format, like pyhmmer. hmmsearch, can then use Python type system to make sure they receive sequences in the right mode. This allows type checkers such as mypy to detect potential contract breaches at compile-time.

checksum()

Calculate a 32-bit checksum for the sequence.

clear()

Reinitialize the sequence for re-use.

accession

The accession of the sequence, if any.

Type str or None

description

The description of the sequence.

 $Type \ {\tt bytes}$

name

The name of the sequence.

Type bytes

source

The source of the sequence, if any.

Type bytes

class pyhmmer.easel.TextSequence(Sequence)

A biological sequence stored in text mode.

Hint: Use the sequence property to access the sequence letters as a Python string.

copy()

Duplicate the text sequence, and return the copy.

digitize()

Convert the text sequence to a digital sequence using alphabet.

sequence

The raw sequence letters, as a Python string.

Type str

class pyhmmer.easel.DigitalSequence(Sequence)

A biological sequence stored in digital mode.

Hint: Use the sequence property to access the sequence digits as a memory view, allowing to access the individual bytes. This can be combined with numpy.asarray to get the sequence as an array with zero-copy.

alphabet

The biological alphabet used to encode this sequence to digits.

Type Alphabet, readonly

copy()

Duplicate the digital sequence, and return the copy.

textize()

Convert the digital sequence to a text sequence.

sequence

The raw sequence digits, as a memory view.

Type memoryview

Sequence File

class pyhmmer.easel.SequenceFile

A wrapper around a sequence file, containing unaligned sequences.

This class supports reading sequences stored in different formats, such as FASTA, GenBank or EMBL. The format of each file can be automatically detected, but it is also possible to pass an explicit format specifier when the *SequenceFile* is instantiated.

guess_alphabet()

Guess the alphabet of an open SequenceFile.

This method tries to guess the alphabet of a sequence file by inspecting the first sequence in the file. It returns the alphabet, or None if the file alphabet cannot be reliably guessed.

Raises

- **EOFError** if the file is empty.
- **OSError** if a parse error occurred.
- ValueError if this methods is called after the file was closed.

read()

Read the next sequence from the file.

Parameters

- **skip_info** (bool) Pass True to disable reading the sequence *metadata*, and only read the sequence *letters*. Defaults to False`.
- **skip_sequence** (bool) Pass True to disable reading the sequence *letters*, and only read the sequence *metadata*. Defaults to False.
- **Returns** Sequence The next sequence in the file, or None if all sequences were read from the file.
- **Raises ValueError** When attempting to read a sequence from a closed file, or when the file could not be parsed.

Hint: This method allocates a new sequence, which is not efficient in case the sequences are being read within a tight loop. Use *SequenceFile.readinto* with an already initialized *Sequence* if you can to recycle the internal buffers.

readinto()

Read the next sequence from the file, using seq to store data.

Parameters

- **seq** (Sequence) A sequence object to use to store the next entry in the file. If this sequence was used before, it must be properly reset (using the Sequence.clear method) before using it again with readinto.
- **skip_info** (bool) Pass True to disable reading the sequence *metadata*, and only read the sequence *letters*. Defaults to False`.
- **skip_sequence** (bool) Pass True to disable reading the sequence *letters*, and only read the sequence *metadata*. Defaults to False.
- **Returns** Sequence A reference to seq that was passed as an input, or None if no sequences are left in the file.

Raises ValueError – When attempting to read a sequence from a closed file, or when the file could not be parsed.

Example

Use SequenceFile.readinto to loop over the sequences in a file while recycling the same Sequence buffer:

```
>>> with SequenceFile("vendor/hmmer/testsuite/ecori.fa") as sf:
... seq = TextSequence()
... while sf.readinto(seq) is not None:
... # ... process seq here ... #
... seq.clear()
```

set_digital()

Set the SequenceFile to read in digital mode with alphabet.

This method can be called even after the first sequences have been read; it only affects subsequent sequences in the file.

Sequence / Subsequence Index

class pyhmmer.easel.SSIReader

A read-only handler for sequence/subsequence index file.

class Entry (fd, record_offset, data_offset, record_length)

property data_offset Alias for field number 2

property fd Alias for field number 0

property record_length Alias for field number 3

property record_offset Alias for field number 1

class FileInfo(name, format)

property format Alias for field number 1

property name Alias for field number 0

class pyhmmer.easel.SSIWriter

A writer for sequence/subsequence index files.

add_file()

Add a new file to the index.

Parameters

- **filename** (*str*) The name of the file to register.
- format (int) A format code to associate with the file, or 0.

Returns int – The filehandle associated with the new indexed file.

2.3.3 Plan7

High-level interface to the Plan7 data model.

Plan7 is the model architecture used by HMMER since HMMER2.

See also:

Details about the Plan 7 architecture in the HMMER documentation.

Alignment

class pyhmmer.plan7.**Alignment** A single alignment of a sequence to a profile.

Background Model

class pyhmmer.plan7.Background The null background model of HMMER.

Domains

class pyhmmer.plan7.**Domain** A single domain in a query *Hit*.

score The overall score in bits, null-corrected.

Type float

class pyhmmer.plan7.**Domains** A sequence of domains corresponding to a single *Hit*.

Hits

class pyhmmer.plan7.Hit

A high-scoring database hit found by the comparison pipeline.

pre_score

Bit score of the sequence before null2 correction.

Type float

score

Bit score of the sequence with all domains after correction.

Type float

class pyhmmer.plan7.TopHits

A ranked list of top-scoring hits.

TopHits are thresholded using the parameters from the pipeline, and are sorted by key when you obtain them from a *Pipeline* instance:

```
>>> abc = thioesterase.alphabet
>>> hits = Pipeline(abc).search(thioesterase, proteins)
>>> hits.is_sorted()
True
```

Use len to query the number of top hits, and the usual indexing notation to extract a particular hit:

```
>>> len(hits)
1
>>> hits[0].name
b'938293.PRJEB85.HG003687_113'
```

clear()

Free internals to allow reusing for a new pipeline run.

HMM

```
class pyhmmer.plan7.HMM
```

A data structure storing the Plan7 Hidden Markov Model.

write()

Write the HMM to a file handle.

Parameters

- **fh** (io.IOBase) A Python file handle, opened in binary mode (this must be the case even with binary=False, since the C code will emit bytes in either case).
- **binary** (bool) Pass False to emit the file in ASCII mode using the latest supported HMMER format, or True to use the binary HMMER3 format.

t zero()

Set all parameters to zero (including model composition).

М

The length of the model (i.e. the number of nodes).

Type int

accession

The accession of the HMM, if any.

Type bytes or None

description

The description of the HMM, if any.

Type bytes or None

name

The name of the HMM, if any.

Type bytes or None

HMM File

class pyhmmer.plan7.HMMFile

A wrapper around a file (or database), storing serialized HMMs.

close()

Close the HMM file and free resources.

This method has no effect if the file is already closed.

Pipeline

class pyhmmer.plan7.Pipeline

An HMMER3 accelerated sequence/profile comparison pipeline.

${\tt search}\,(\,)$

Run the pipeline using a query HMM against a sequence database.

Parameters

- hmm (HMM) The HMM object to use to query the sequence database.
- **sequences** (Iterable of *DigitalSequence*) The sequences to query with the HMMs. Pass a SequenceFile instance in digital mode to iteratively read from disk.
- hits (*TopHits*, optional) A hit collection to store results in, if any. If None, a new collection will be allocated. Use a non-None value if you are running several HMMs sequentially and want to aggregate the results, or if you are able to recycle a *TopHits* instance with *TopHits.clear*.
- **Returns** *TopHits* the hits found in the sequence database. If an instance was passed via the hits argument, it is safe to ignore the return value and to access it by reference.
- **Raises ValueError** When the alphabet of the current pipeline does not match the alphabet of the given HMM.

Profile

class pyhmmer.plan7.Profile A Plan7 search profile.

A I fail / scatch pro

clear()

Clear internal buffers to reuse the profile without reallocation.

configure()

Configure a search profile using the given models.

Parameters

- hmm (pyhmmer.plan7.HMM) The model HMM with core probabilities.
- **bg** (*pyhmmer.plan7.Background*) The null background model.
- L (int) The expected target sequence length.
- **multihit** (bool) Whether or not to use multihit modes.
- **local** (bool) Whether or not to use non-local modes.

is_local()

Returns whether or not the profile is in a local alignment mode.

```
is_multihit()
```

Returns whether or not the profile is in a multihit alignment mode.

```
optimized()
```

Convert the profile to a platform-specific optimized profile.

```
class pyhmmer.plan7.OptimizedProfile
```

An optimized profile that uses platform-specific instructions.

```
\texttt{is\_local()}
```

Returns whether or not the profile is in a local alignment mode.

```
write()
```

Write an optimized profile to two separate files.

HMMER implements an acceleration pipeline using several scoring algorithms. Parameters for MSV (the *Multi ungapped Segment Viterbi*) are saved independently to the fh_filter handle, while the rest of the profile is saved to fh_profile.

Cython bindings and Python interface to HMMER3.

HMMER is a biological sequence analysis tool that uses profile hidden Markov models to search for sequence homologs. HMMER3 is maintained by members of the the Eddy/Rivas Laboratory at Harvard University.

pyhmmer is a module, implemented using the Cython language, that provides bindings to HMMER3. It directly interacts with the HMMER internals, which has several advantages over CLI wrappers like hmmer-py.

2.4 Contributing to pyHMMER

For bug fixes or new features, please file an issue before submitting a pull request. If the change isn't trivial, it may be best to wait for feedback.

2.4.1 Setting up a local repository

Make sure you clone the repository in recursive mode, so you also get the wrapped code of Easel and HMMER, which are exposed as git submodules:

\$ git clone --recursive https://github.com/althonos/pyhmmer

2.4.2 Running tests

Tests are written as usual Python unit tests with the unittest module of the standard library. Running them requires the extension to be built locally:

```
$ python setup.py build_ext --debug --inplace
$ python -m unittest discover -vv
```

2.4.3 Coding guidelines

This project targets Python 3.6 or later.

Python objects should be typed; since it is not supported by Cython, you must manually declare types in type stubs (.pyi files). In Python files, you can add type annotations to function signatures (supported in Python 3.5) or in variable assignments (supported from Python 3.6 onward).

Interfacing with C

When interfacing with C, and in particular with pointers, use assertions everywhere you assume the pointer to be non-NULL.

2.5 Changelog

All notable changes to this project will be documented in this file.

The format is based on Keep a Changelog and this project adheres to Semantic Versioning.

2.5.1 Unreleased

2.5.2 v0.1.4 - 2021-01-15

Added

- DigitalSequence.textize to convert a digital sequence to a text sequence.
- DigitalSequence.___init___ method allowing to create a digital sequence from any object implementing the buffer protocol.
- Alignment.hmm_accession property to retrieve the accession of the HMM in an alignment.

2.5.3 v0.1.3 - 2021-01-08

Fixed

• Compilation issues in OSX-specific Cython code.

2.5.4 v0.1.2 - 2021-01-07

Fixed

• Required Cython files not being included in source distribution.

2.5.5 v0.1.1 - 2020-12-02

Fixed

- HMMFile calling file.peek without arguments, causing it to crash when passed some types, e.g. gzip. GzipFile.
- HMMFile failing to work with PyPy file objects because of a bug with their implementation of readinto.
- C/Python file object implementation using stropy instead of memopy, causing issues when null bytes were read.

2.5.6 v0.1.0 - 2020-12-01

Initial beta release.

Fixed

- TextSequence uses the sequence argument it's given on instantiation.
- Segmentation fault in Sequence. ____eq___ caused by implicit type conversion.
- Segmentation fault on SequenceFile.read failure.
- Missing type annotations for the pyhmmer.easel module.

2.5.7 v0.1.0-a5 - 2020-11-28

Added

- Sequence.__len__ magic method so that len (seq) returns the number of letters in seq.
- Python file-handle support when opening an pyhmmer.plan7.HMMFile.
- Context manager protocol to pyhmmer.easel.SSIWriter.
- Type annotations for pyhmmer.easel.SSIWriter.
- add_alias to pyhmmer.easel.SSIWriter.
- write method to pyhmmer.plan7.OptimizedProfile to write an optimized profile in binary format.
- offsets property to interact with the disk offsets of a pyhmmer.plan7.OptimizedProfile instance.
- pyhmmer.hmmer.hmmpress emulating the hmmpress binary from HMMER.
- M property to pyhmmer.plan7.HMM exposing the number of nodes in the model.

Changed

- Bumped vendored Easel to v0.48.
- Bumped vendored HMMER to v3.3.2.
- pyhmmer.plan7.HMMFile will raise an EOFError when given an empty file.
- Renamed length property to L in pyhmmer.plan7.Background.

Fixed

- Segmentation fault when close method of pyhmmer.easel.SSIWriter was called more than once.
- close method of pyhmmer.easel.SSIWriter not writing the index contents.

2.5.8 v0.1.0-a4 - 2020-11-24

Added

- MSA, TextMSA and DigitalMSA classes representing a multiple sequence alignment to pyhmmer.easel.
- Methods and protocol to copy a Sequence and a MSA.
- pyhmmer.plan7.OptimizedProfile wrapping a platform-specific optimized profile.
- SSIReader and SSIWriter classes interacting with sequence/subsequence indices to pyhmmer.easel.
- Exception handler using Python exceptions to report Easel errors.

Changed

- pyhmmer.hmmsearch returns an iterator of TopHits, with one instance per HMM in the input.
- pyhmmer.hmmsearch properly raises errors happenning in the background threads without deadlock.
- pyhmmer.plan7.Pipeline recycles memory between Pipeline.search calls.

Fixed

• Missing type annotations for the pyhmmer.errors module.

Removed

• Unneeded or private methods from pyhmmer.plan7.

2.5.9 v0.1.0-a3 - 2020-11-19

Added

- TextSequence and DigitalSequence representing a Sequence in a given mode.
- E-value properties to Hit and Domain.
- TopHits now stores a reference to the pipeline it was obtained from.
- Pipeline.Z and Pipeline.domZ properties.
- Experimental pickling support to Alphabet.
- Experimental freelist to Sequence class to avoid allocation bottlenecks when iterating on a SequenceFile without recycling sequence buffers.

Changed

- Made Sequence an abstract base class.
- Additional Pipeline parameters can be passed as keyword arguments to pyhmmer.hmmsearch.
- SequenceFile.read can now be configured to skip reading the metadata or the content of a sequence.

Removed

• Redundant SequenceFile methods.

Fixed

- doctest loader crashing on Python 3.5.
- TopHits.threshold segfaulting when being called without prior Tophits.sort call
- Unknown format argument to SequenceFile constructor not raising the right error.

2.5.10 v0.1.0-a2 - 2020-11-12

Added

- Support for compilation on PowerPC big-endian platforms.
- Type annotations and stub files for Cython modules.

Changed

- distutils is now used to compile the package, instead of calling autotools and letting HMMER configure itself.
- Bitfield.count now allows passing an argument (for compatibility with collections.abc. Sequence).

2.5.11 v0.1.0-a1 - 2020-11-10

Initial alpha release (test deployment to PyPI).

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